



Enhanced Virus Detection and Metagenomic Sequencing in Patients with Meningitis and Encephalitis

 Anne Piantadosi,^{a,b,c} Shibani S. Mukerji,^{d,e} Simon Ye,^{a,f} Michael J. Leone,^d Lisa M. Freimark,^a Daniel Park,^a Gordon Adams,^{a,b} Jacob Lemieux,^{a,b,e}  Sanjat Kanjilal,^{b,e,g} Isaac H. Solomon,^{e,g} Asim A. Ahmed,^{e,h} Robert Goldstein,^{b,e} Vijay Ganesh,^{a,e,i} Bridget Ostrem,^{d,e} Kaelyn C. Cummins,^{j*} Jesse M. Thon,^{d,e*} Cormac M. Kinsella,^{a,m*} Eric Rosenberg,^{b,e,k} Matthew P. Frosch,^{d,e,k} Marcia B. Goldberg,^{b,e} Tracey A. Cho,^{d,e,l} Pardis Sabeti^{a,b,e,m,n,o}

^aBroad Institute of MIT and Harvard, Cambridge, Massachusetts, USA

^bDivision of Infectious Diseases, Massachusetts General Hospital, Boston, Massachusetts, USA

^cEmory University School of Medicine, Atlanta, Georgia, USA

^dDepartment of Neurology, Massachusetts General Hospital, Boston, Massachusetts, USA

^eHarvard Medical School, Boston, Massachusetts, USA

^fHarvard-MIT Program of Health Sciences and Technology, Cambridge, Massachusetts, USA

^gDepartment of Pathology, Brigham and Women's Hospital, Boston, Massachusetts, USA

^hDepartment of Pediatrics, Harvard Medical School, Children's Hospital, Boston, Massachusetts, USA

ⁱDepartment of Neurology, Brigham and Women's Hospital, Boston, Massachusetts, USA

^jDivision of Infectious Diseases, Brigham and Women's Hospital, Boston, Massachusetts, USA

^kDepartment of Pathology, Massachusetts General Hospital, Boston, Massachusetts, USA

^lUniversity of Iowa, Department of Neurology, Iowa City, Iowa, USA

^mDepartment of Organismic and Evolutionary Biology, Harvard University, Cambridge, Massachusetts, USA

ⁿDepartment of Immunology and Infectious Disease, Harvard T. H. Chan School of Public Health, Boston, Massachusetts, USA

^oHoward Hughes Medical Institute, Chevy Chase, Maryland, USA

Anne Piantadosi, Shibani S. Mukerji, and Simon Ye all contributed equally to this work. Author order was determined by mutual agreement among first authors. Tracey A. Cho and Pardis Sabeti contributed equally.

ABSTRACT Meningitis and encephalitis are leading causes of central nervous system (CNS) disease and often result in severe neurological compromise or death. Traditional diagnostic workflows largely rely on pathogen-specific tests, sometimes over days to weeks, whereas metagenomic next-generation sequencing (mNGS) profiles all nucleic acid in a sample. In this single-center, prospective study, 68 hospitalized patients with known ($n=44$) or suspected ($n=24$) CNS infections underwent mNGS from RNA and DNA to identify potential pathogens and also targeted sequencing of viruses using hybrid capture. Using a computational metagenomic classification pipeline based on KrakenUniq and BLAST, we detected pathogen nucleic acid in cerebrospinal fluid (CSF) from 22 subjects, 3 of whom had no clinical diagnosis by routine workup. Among subjects diagnosed with infection by serology and/or peripheral samples, we demonstrated the utility of mNGS to detect pathogen nucleic acid in CSF, importantly for the *Ixodes scapularis* tick-borne pathogens Powassan virus, *Borrelia burgdorferi*, and *Anaplasma phagocytophilum*. We also evaluated two methods to enhance the detection of viral nucleic acid, hybrid capture and methylated DNA depletion. Hybrid capture nearly universally increased viral read recovery. Although results for methylated DNA depletion were mixed, it allowed the detection of varicella-zoster virus DNA in two samples that were negative by standard mNGS. Overall, mNGS is a promising approach that can test for multiple pathogens simultaneously, with efficacy similar to that of pathogen-specific tests, and can uncover geographically relevant infectious CNS disease, such as tick-borne infections in New England. With further laboratory and computational enhancements, mNGS may become a mainstay of workup for encephalitis and meningitis.

Citation Piantadosi A, Mukerji SS, Ye S, Leone MJ, Freimark LM, Park D, Adams G, Lemieux J, Kanjilal S, Solomon IH, Ahmed AA, Goldstein R, Ganesh V, Ostrem B, Cummins KC, Thon JM, Kinsella CM, Rosenberg E, Frosch MP, Goldberg MB, Cho TA, Sabeti P. 2021. Enhanced virus detection and metagenomic sequencing in patients with meningitis and encephalitis. mBio 12:e01143-21. <https://doi.org/10.1128/mBio.01143-21>.

Editor Nisha Duggal, Virginia Polytechnic Institute and State University

Copyright © 2021 Piantadosi et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Anne Piantadosi, anne.piantadosi@emory.edu, or Shibani S. Mukerji, smukerji@partners.org.

* Present address: Kaelyn C. Cummins, Baylor College of Medicine, Houston, Texas, USA; Jesse M. Thon, Department of Neurology, University of Pennsylvania, Philadelphia, Pennsylvania, USA; Cormac M. Kinsella, Amsterdam UMC, Amsterdam, The Netherlands.

Received 4 May 2021

Accepted 2 July 2021

Published 31 August 2021

IMPORTANCE Meningitis and encephalitis are leading global causes of central nervous system (CNS) disability and mortality. Current diagnostic workflows remain inefficient, requiring costly pathogen-specific assays and sometimes invasive surgical procedures. Despite intensive diagnostic efforts, 40 to 60% of people with meningitis or encephalitis have no clear cause of CNS disease identified. As diagnostic uncertainty often leads to costly inappropriate therapies, the need for novel pathogen detection methods is paramount. Metagenomic next-generation sequencing (mNGS) offers the unique opportunity to circumvent these challenges using unbiased laboratory and computational methods. Here, we performed comprehensive mNGS from 68 prospectively enrolled patients with known ($n=44$) or suspected ($n=24$) CNS viral infection from a single center in New England and evaluated enhanced methods to improve the detection of CNS pathogens, including those not traditionally identified in the CNS by nucleic acid detection. Overall, our work helps elucidate how mNGS can become integrated into the diagnostic toolkit for CNS infections.

KEYWORDS encephalitis, metagenomic sequencing, next-generation sequencing (NGS), meningitis, virus, hybrid capture, methylated DNA depletion

Meningitis and encephalitis are leading causes of central nervous system (CNS) disease, ranked as the 4th leading contributor to global neurological disability-adjusted life-years (1), often resulting in severe neurological compromise or death (2, 3). Traditional diagnostic workflows remain inefficient, requiring costly pathogen-specific diagnostics, serial cerebrospinal fluid (CSF) testing, and sometimes invasive surgical procedures. Despite these intensive diagnostic efforts, 40 to 60% of subjects with meningitis or encephalitis have no clear cause identified (2, 4–6).

Metagenomic next-generation sequencing (mNGS) offers a unique opportunity to circumvent some of these challenges. mNGS consists of unbiased sequencing of all nucleic acid in a sample followed by computational classification of reads to identify potential pathogens (7–9). This technique has successfully detected a range of pathogens, including bacteria (10–12), fungi (13), protozoa (14), and viruses (15–17), in subjects with CNS infection. mNGS is increasingly used as a clinical diagnostic test (18–20), and criteria for test performance have been described but not yet standardized (21–23).

In this study, we prospectively enrolled 68 patients with known or suspected CNS infection and performed mNGS from both RNA and DNA to identify pathogens. We focused laboratory and analysis methods on viral nucleic acid detection since viruses are the most common type of pathogen detected in CNS infection (4, 5, 24, 25). The goals of this study were to assess the utility of standard mNGS in identifying CNS pathogens and to examine enhanced laboratory techniques for improving analytical sensitivity, including hybrid capture (HC) of viral nucleic acid and methylated DNA depletion (MDD).

RESULTS

Clinical characteristics. Of the 68 adults enrolled, 63% (43/68) were male, subjects ranged in age from 24 to 86 years (median = 58 years [interquartile range {IQR}, 39, 72 years]) (Table 1), and 25 (37%) were immunocompromised (Fig. 1; see also Text S1 in the supplemental material). New England was the primary residence for all except one subject, who lived in Florida. Altered mental status was described in 56% (38/68), while a minority had photophobia (24% [16/68]) or neck stiffness (26% [18/68]). Twenty subjects out of 68 (29%) were admitted to the intensive care unit (ICU), and the in-hospital mortality rate was 6% (4/68).

Based on clinical testing, 44 of the 68 subjects received a conclusive diagnosis by discharge. Twelve subjects were diagnosed with viral infection by PCR from CSF (“infection, CSF PCR+” group), 25 were diagnosed with infection by serology or PCR from blood (“infection, other” group), and 7 had a noninfectious etiology (“alternative diagnosis” group). The remaining 24 subjects (35%) had no known diagnosis (“unknown” group) (Table 1). Subjects classified as “unknown” underwent exhaustive clinical testing; 50% of them (12/24) had ≥ 25 infectious disease (ID) tests (Fig. 2A; Fig. S2 and Table S2), and no diagnoses were

TABLE 1 Clinical characteristics of enrolled subjects stratified by diagnostic group^a

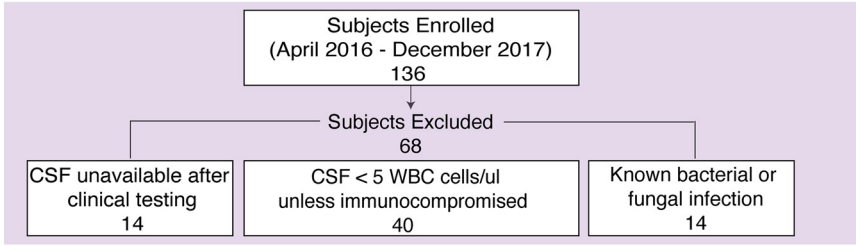
Characteristic	Value for group				
	Overall (n = 68)	Infection, PCR ⁺ (n = 12)	Infection, other (n = 25)	Alternative diagnosis (n = 7)	Unknown (n = 24)
Demographics					
Median age (yrs) (IQR)	58.5 (39, 72.3)	57.5 (39, 67.3)	61 (43, 72)	73 (37.5, 77)	57.5 (38, 71)
No. of male subjects (%)	43 (63)	5 (42)	19 (76)	6 (86)	13 (54)
No. of immunocompetent subjects (%)	43 (63)	7 (58)	17 (68)	5 (71)	14 (58)
Median length of stay (days) (min, max)	8 (2, 51)	4.5 (2, 51)	9 (3, 51)	12 (3, 30)	7 (5, 22)
No. of subjects of race (%)					
White	57 (84)	10 (83.3)	21 (84)	6 (85.7)	20 (83.3)
Black or African American	2 (3)	1 (8)	0 (0)	1 (14)	0 (0)
No. of subjects with time from symptom onset to LP (%)					
Acute (0–3 days)	17 (25)	2 (16.7)	7 (28)	1 (14.3)	7 (29.2)
Early subacute (4–7 days)	12 (17.6)	5 (41.7)	3 (12)	1 (14.3)	3 (12.5)
Late subacute (8–30 days)	30 (44.1)	3 (25)	14 (56)	2 (28.6)	11 (45.8)
Chronic (>30 days)	9 (13.2)	2 (16.7)	1 (4)	3 (42.9)	3 (12.5)
Symptoms and signs during hospitalization					
No. of subjects with altered mental status (%)	38 (56)	7 (58.3)	14 (56)	5 (71.4)	12 (50)
No. of subjects with photophobia (%)	16 (24)	3 (25)	4 (16)	2 (28.6)	7 (29.2)
No. of subjects with neck stiffness (%)	18 (27)	2 (16.7)	6 (24)	2 (28.6)	8 (33.3)
Median max temp (°C) (IQR)	38.1 (37.4, 39)	37.8 (37.4, 38.1)	38.1 (37.6, 38.9)	37.7 (37.6, 39)	38.4 (37.4, 39.2)
No. of subjects with fever (max ≥ 38°C) (%)	38 (56)	6 (50)	14 (56)	3 (42.9)	15 (62.5)
Laboratory data					
Median laboratory parameter value (IQR)					
Hematology					
White blood cell count (WBCs/ μ l)	8.6 (7.4, 10.2)	8.8 (8.1, 9.6)	7.84 (6.5, 9.3)	8.7 (8.2, 9.7)	9.6 (7.5, 10.9)
CSF					
White blood cell count (WBCs/ μ l)	80.5 (16.8, 131.5)	105.5 (35.5, 337)	47 (14, 105)	17 (10, 25.5)	98.5 (40.5, 133.5)
Total protein (mg/dl)	70.5 (50, 117)	51.5 (39.8, 111)	65 (55, 117)	69 (44.5, 92.5)	78.5 (55, 120)
Glucose (mg/dl)	62 (54, 73.5)	67.5 (55, 82)	62 (55, 69)	60 (55, 64)	60 (52, 73.5)
Median no. of infectious disease tests ordered (min, max)	19 (6, 62)	12 (6, 56)	25 (6, 62)	26 (10, 57)	22.5 (6, 48)
No. of subjects with admission service (%)					
Medicine floor/ICU	21 (31)	4 (33.3)	9 (36)	0 (0)	8 (33.3)
Neurology floor/ICU	37 (54)	6 (50)	13 (52)	7 (100)	11 (45.8)
Other	10 (15)	2 (16.7)	3 (12)	0 (0)	5 (20.8)
Admission to ICU during hospitalization	20 (29)	3 (25)	9 (36)	3 (42.9)	5 (20.8)
No. of subjects admitted during study period (%)					
1 December–28 February	11 (16)	2 (16.7)	4 (16)	1 (14.3)	4 (16.7)
1 March–31 May	14 (21)	3 (25)	5 (20)	2 (28.6)	4 (16.7)
1 June–31 August	19 (28)	3 (25)	4 (16)	3 (42.9)	9 (37.5)
1 September–30 November	24 (35)	4 (33.3)	12 (48)	1 (14.3)	7 (29.2)
No. of subjects with postdischarge outcome (%)					
Home	39 (57)	5 (41.7)	12 (48)	3 (42.9)	19 (79.2)
Rehabilitation ^b	25 (37)	5 (41.7)	12 (48)	3 (42.9)	5 (20.8)
Death	4 (6)	2 (16.7)	1 (4)	1 (14.3)	0 (0)

^aAbbreviations: CSF, cerebrospinal fluid; IQR, interquartile range; PCR⁺, positive PCR; ICU, intensive care unit.

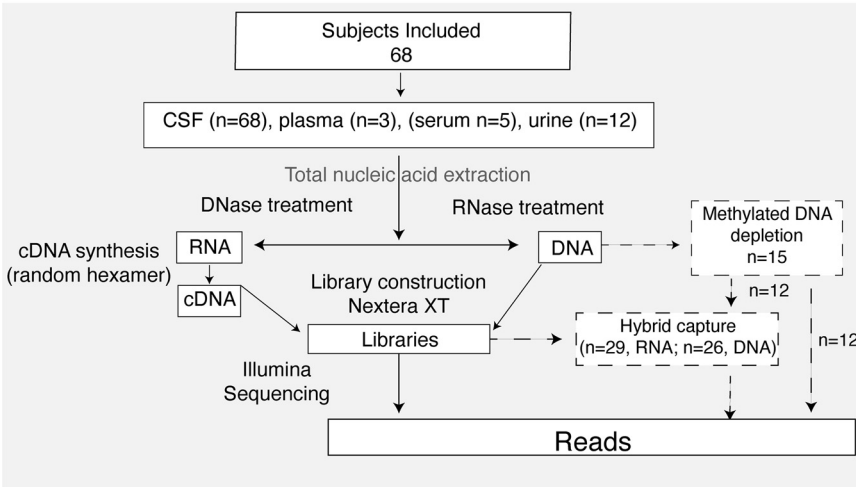
^bLong-term acute care or skilled nursing facility.

made during long-term follow-up (Table S3). In contrast, the “infection, CSF PCR⁺” group had a much lower median number of clinical ID tests performed (12 [IQR, 6, 56] versus 22.5 [IQR, 11, 36] for the “unknown” group). The “infection, CSF PCR⁺” group also had the shortest length of stay (LOS) (4.5 days [IQR, 2, 51 days]), and across the total cohort, the LOS moderately correlated with the number of ID tests ordered (Spearman’s $\rho = 0.65$; $P < 0.01$) (Fig. 2B).

A. Enrollment



B. Laboratory methods



C. Analyses methods

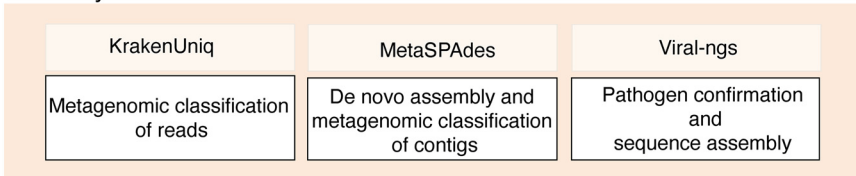


FIG 1 Overview of methods for subject selection and mNGS. Enrollment (A), laboratory methods (B), and analysis methods (C) are shown. Enhanced laboratory methods for methylated DNA depletion and hybrid capture (dashed lines) were included for a subset of the samples as shown. Abbreviations: CSF, cerebrospinal fluid; WBC, white blood cell.

Results from mNGS and enhanced methods. To understand mNGS performance in a real-world context, we sequenced 68 CSF, 3 plasma, 5 serum, and 12 urine samples along with 47 negative controls. We performed mNGS from RNA, DNA, or both, generating an average of 9.6 million reads per subject (Fig. S3; see also Tables S4 to S6 at <https://figshare.com/articles/dataset/Tables/13266506>). We identified a plausible pathogen in 22 subjects (32.4%): 18 by standard mNGS, an additional 2 with the use of HC, and 2 more with the use of MDD (Fig. 3; Fig. S4; see also Table S7 at <https://figshare.com/articles/dataset/Tables/13266506>). As expected, we detected viral nucleic acid in most subjects in the “infection, CSF PCR+” group (10 out of 12 [83%]) (Fig. 3), consistent with other mNGS studies (18, 23). mNGS was negative in one subject with herpes simplex virus 2 (HSV-2) infection and another with human immunodeficiency virus type 1 (HIV-1) infection, illustrating that mNGS can be less sensitive than PCR for very low-level infections (Text S1 and Fig. S5). We detected reads from both JC virus and HIV in a subject with HIV and progressive multifocal leukoencephalopathy (PML), illustrating the capacity of this single platform to identify viral coinfections. In assessing our enhanced methods, we found that HC increased the number of viral reads in 8 out of 9 cases positive by routine mNGS, sometimes substantially (Fig. 4). In contrast, MDD led to mixed results, enabling pathogen detection in 4 out of 19 matched libraries (e.g., varicella-zoster virus [VZV] in subjects M049 and M070) and enhanced yields in 5 additional libraries while decreasing the yields in 10 libraries (e.g., Epstein-Barr virus [EBV] in subject M095) (Fig. 4; Text S1).

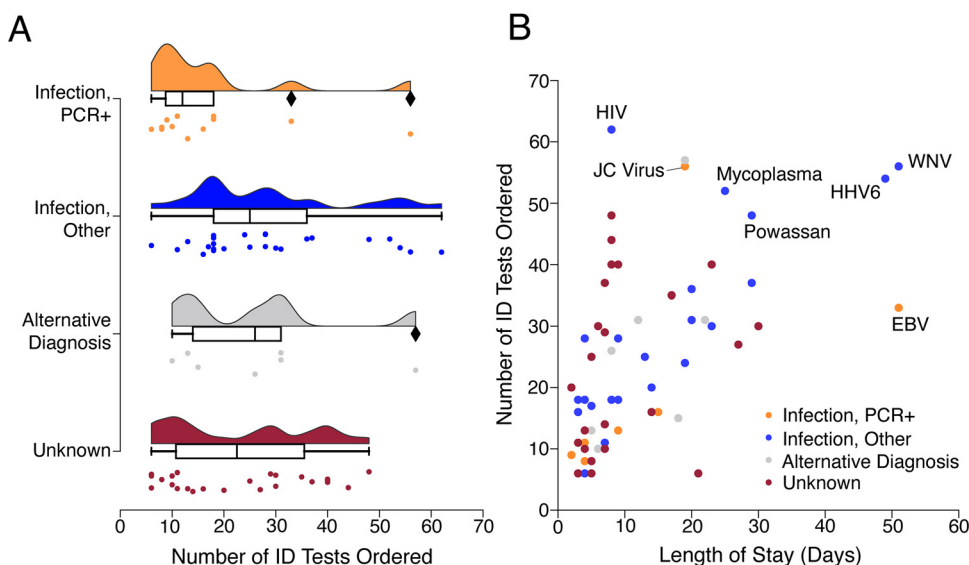


FIG 2 Number of infectious disease tests ordered and lengths of stay among subjects. (A) Distributions showing the number of infectious disease (ID)-related tests ordered per subject, stratified by clinical diagnosis category. ID tests were counted if ordered between hospital admission day 1 and hospital discharge. Box plots with horizontal bars represent medians and interquartile ranges for ID tests. Diamonds represent data points greater than $1.5 \times$ the IQR. (B) Scatterplot showing the number of ID tests versus length of stay per subject. Colors indicate clinical diagnosis categories. The LOS correlated with the number of total ID tests ordered (Spearman's $\rho = 0.65$; $P < 0.01$). The final clinical diagnosis for viral pathogens is stated for cases whose number of ID tests or LOS was an outlier above the 3rd quartile.

mNGS and enhanced methods detect pathogens not traditionally detected by CSF PCR. Twenty-five subjects in the “infection, other” group had infections diagnosed by serology from CSF and/or blood or by PCR from blood (Table 1). Fifteen had an infection for which no clinically approved CSF PCR assay was available; standard mNGS detected pathogen nucleic acid in six, and mNGS plus HC detected pathogen nucleic acid in a seventh, yielding 7/15 positive hits (47%) (Fig. 3). There were several cases of regional interest. For example, three subjects were clinically diagnosed with Powassan encephalitis using a time-consuming send-out serology test, and mNGS identified Powassan virus RNA in two cases. In addition, while our methods were focused on viral detection, we identified atypical bacteria whose genome reads were readily distinguishable from the background, including *Borrelia burgdorferi* in two out of two subjects diagnosed with Lyme disease by serology and *Anaplasma phagocytophilum* in a subject diagnosed by PCR from blood (Fig. S6).

In the remaining 10 subjects from the “infection, other” group, clinical CSF PCR was available and negative for the culprit pathogen (human herpesvirus 6 [HHV-6] [$n = 1$], VZV [$n = 3$], West Nile virus [WNV] [$n = 3$], and mycoplasma [$n = 3$]). While all of these were negative using standard mNGS, the addition of MDD allowed the detection of VZV in two subjects (M049 and M070) (Text S1; see also Fig. S7 at https://figshare.com/articles/figure/Supplemental_Figures/13266488). In both cases, clinical CSF VZV PCR from the same sample was negative, illustrating that mNGS may occasionally be more sensitive than a clinically validated PCR. In contrast, MDD decreased the yields for other herpesviruses, suggesting pathogen-specific effects (Fig. 4; Text S1).

mNGS detects pathogens not tested by clinicians. Among the 24 subjects with no identified clinical diagnosis (“unknown”), standard mNGS identified viruses in 3 subjects, and no additional pathogens were detected using MDD and HC. We detected enterovirus in two subjects with lymphocytic meningitis (subjects M108 and M126), neither of whom had orders for clinical enterovirus PCR. These findings were verified by sequencing a second CSF aliquot and by assembling a complete enterovirus genome for each subject. Phylogenetic analysis for both subjects demonstrated closely related echovirus 30 strains (see Fig. S8 at https://figshare.com/articles/figure/Supplemental_Figures/13266488).

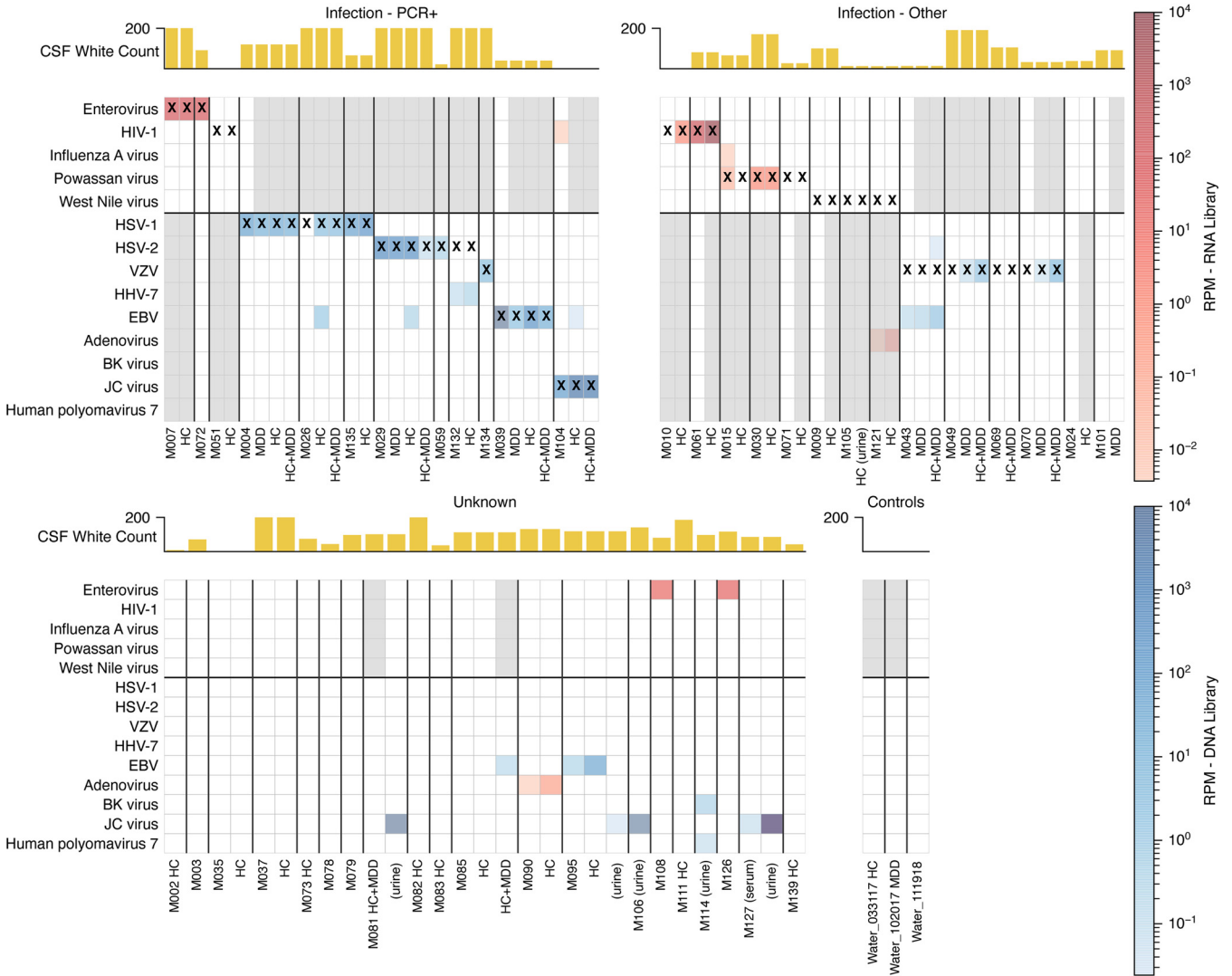


FIG 3 Viral taxa identified in cerebrospinal fluid using mNGS with or without enhanced methods. A heat map shows viral taxa identified in each sample. Rows are viral taxa, and columns are samples, some with enhanced sequencing methods (HC and/or MDD). Only classifications with over 100 unique kmers, with at least 1 BLAST-confirmed read, and manually reviewed as noncontaminants are shown. Rows are grouped by RNA viruses (top section) or DNA viruses (bottom section). The color intensity corresponds to the RPM of the taxa. Red boxes correspond to detection in RNA libraries, while blue boxes correspond to detection in DNA libraries. Some DNA viruses were detected in RNA libraries (e.g., adenovirus in subject M121). Gray-shaded columns represent samples that did not undergo DNA or RNA sequencing. Samples in which a contaminant was found are included here as blank columns, and the contaminants are shown in Fig. S4 in the supplemental material. X's represent the clinical diagnosis. Yellow bars indicate the CSF nucleated cell count for each subject. The four groupings of columns from the top left to the bottom right correspond to infections diagnosed by positive PCR, infections diagnosed by nonmolecular techniques, subjects with an unknown etiology, and negative controls, including extracted water.

We also detected EBV and assembled a complete genome in one subject (M095) during two serial hospitalizations for recurrent lymphocytic meningitis. While clinical testing for EBV in CSF was not performed, EBV PCR was positive from blood during both admissions. Overall, these results are compatible with EBV meningitis or reactivation in the setting of another, unidentified primary syndrome (26).

mNGS and enhanced methods detect viruses of uncertain significance. In addition to the plausible pathogens described above, we detected DNA viruses of uncertain clinical significance. EBV was present at low levels in CSF from four subjects, three of whom had alternative primary diagnoses: VZV (subject M043), HSV-1 (M026), and HSV-2 (M029). For the fourth subject (M085), no alternative diagnosis was identified; however, EBV reads were detected only after MDD and HC, and a clinical PCR for EBV from CSF was negative. A review of clinical data for these subjects suggested that EBV was unlikely to explain their clinical syndromes, and these findings most likely suggest reactivation in the setting of another acute process.

We also detected human herpesvirus 7 (HHV-7) at a low level in a subject (M132) who was diagnosed with HSV-2 by clinical PCR, but HSV-2 was not detected by mNGS. Acute encephalitis due to HHV-7 rarely occurs in immunocompetent adults and has been described in three cases of patients with limbic encephalitis (27), facial cranial palsy, and polymyeloradiculitis (28, 29); none of these syndromes were compatible with this subject's presentation. Adenovirus reads were detected in two subjects (M090 and M121) and were not considered vector contaminants due to their distribution across the genome; however, the reads were found in RNA libraries only, and subjects were not known to be immunocompromised or to have features compatible with adenovirus infection.

A known challenge of mNGS is the assessment and interpretation of background contamination. Even after extensive computational depletion of both human reads and sequences found in negative controls, bacteria accounted for ~11% of DNA and ~39% of RNA reads. We also found viral reads from bacteriophages and vectors commonly used in molecular biology, such as adenovirus, cytomegalovirus (CMV), HIV/lentiviruses, and parvoviruses, consistent with previous studies (30). Finally, we found a few reads matching recently discovered picornaviruses from environmental surveys (Text S1; see also Table S8 at <https://figshare.com/articles/dataset/Tables/13266506>) (31).

mNGS is negative for subjects with noninfectious diagnoses. mNGS did not detect pathogen nucleic acid in the seven subjects with noninfectious diseases ("alternative diagnosis" group): autoimmune encephalitis and cerebellitis ($n=3$), lymphoma ($n=2$), and vasculitis ($n=2$). In this category, the median CSF white blood cell (WBC) count was 2 to 6 times lower than those in the two infection groups. The "alternative diagnosis" group had the highest number of ID tests ordered for CSF and blood (median, 26 tests [range, 10, 57]) (see Table S2 at <https://figshare.com/articles/dataset/Tables/13266506>), which is consistent with provider practice to test a wide range of pathogens prior to immunomodulatory therapy; subjects were ultimately treated with immunosuppressive agents.

DISCUSSION

Advances in genomic technologies provide translational researchers the unprecedented capacity to identify and study pathogens in patients with meningitis and encephalitis. Here, we performed a prospective study using mNGS, enhanced laboratory and analysis techniques, and detailed clinical phenotyping to assess the use of this technology as a diagnostic tool for hospitalized subjects with inflammatory CSF. We identified a range of CNS pathogens, including regionally important tick-borne organisms not typically detected by CSF nucleic acid testing. In 9 cases, we were able to recover full or partial viral genomes, demonstrating the utility of this technique for virus characterization studies (e.g., molecular epidemiology and identification of neurotropic variants). In our study, subjects with CNS infections diagnosed using CSF PCR underwent fewer ID tests than other clinical groups with inflammatory CSF and had shorter lengths of hospital stay (32); from this, we infer that the judicious application of molecular diagnostic techniques such as mNGS can positively impact patient care and associated costs. Together with recent reports (18), this work highlights the opportunity for mNGS to become integrated into the infectious disease diagnostic toolkit.

Overall, mNGS was highly effective at detecting pathogens identified by clinical PCR. mNGS detected the expected pathogen in 10 of 12 subjects, similar to a recent study detecting viruses in 14 out of 16 subjects diagnosed by CSF PCR (18). Our results also highlight the benefit of enhanced mNGS techniques. Panviral HC consistently improved the sequencing of RNA and DNA viruses and resulted in virus detection in two cases (HSV-1 and HIV) that were negative by standard mNGS. MDD plus mNGS detected VZV DNA in two samples negative by standard mNGS. However, MDD decreased the yields for some viruses, indicating that the role of this technique in mNGS remains unclear. Our mixed results with MDD correlate with previous studies focusing on bacterial metagenomics (33, 34). Saponin lysis of host cells may prove a more effective depletion technique for DNA (35, 36).

A notable strength of this study was the detection of pathogens not routinely detected by CSF PCR, most notably the tick-borne pathogens Powassan virus (17), *Borrelia burgdorferi*, and *Anaplasma phagocytophilum*. These pathogens show increasing rates of human infection (37), particularly in the Northeastern United States, where this study was conducted. For Powassan virus, which is routinely detected by serology, our findings illustrate the potential utility of nucleic acid-based screening. Interestingly, we detected the CSF presence of *Anaplasma*, which is not commonly considered to be a cause of CNS infection (38), although the related intracellular bacterium *Ehrlichia chaffeensis* can cause meningoencephalitis (39, 40). Overall, the high number of subjects with tick-borne infection highlights the importance of conducting mNGS in diverse geographical regions for both diagnostic purposes and epidemiological studies.

Among the 24 subjects for whom no diagnosis was achieved by routine clinical testing (“unknown”), mNGS detected potential pathogens in 3 (8%), a rate similar to that reported previously by Wilson et al. (13/159 [8%]) (18). It is possible that subjects in whom no pathogen nucleic acid was detected had a noninfectious syndrome or an infection with a low pathogen burden or short duration of replication. We reviewed the postdischarge clinical course in the subgroup, and none were identified as having an infectious syndrome, signaling the likelihood that mNGS did not miss an actionable result.

Our results highlight a few challenges associated with mNGS, particularly for infections with low titers or parainfectious complications. For example, we report an equivocal mNGS result in a subject with HIV-1 who had a CSF HIV load of 469 copies/ml, a value close to the recently reported CSF limit of detection of 313 copies/ml for HIV-1 using mNGS (23). Additionally, mNGS results were negative in all four subjects with WNV, three of whom had clinical WNV PCRs from CSF performed, which were also negative. These results support other studies showing that WNV nucleic acid is usually undetectable in CSF by clinical PCR (23, 41) or mNGS (18), although it may be observed in immunocompromised subjects (15, 41–44). Similarly, CSF mycoplasma nucleic acid was not detected clinically or by mNGS in three subjects despite positive mycoplasma IgM serologies. These patients had clinical and neuroradiographic findings suggestive of encephalitis or encephalomyelitis, including multifocal T2/FLAIR (T2-weighted fluid-attenuated inversion recovery) hyperintensities in the brain and spinal cord (M006), superrefractory epilepsy with T2/FLAIR hyperintensity in the right posterior subinsular/anterior temporal stem (M032), and longitudinally extensive transverse myelitis (M075). This finding supports interpretations that CNS complications of mycoplasma infections likely reflect a parainfectious antibody-mediated response rather than direct infection (45).

While we investigated specific atypical bacteria of interest (*Borrelia*, *Anaplasma*, and *Mycoplasma* spp.), our study focused on viruses for four key reasons: they are the most common pathogens in CNS infection (4, 5, 24, 25); bacteria and fungi often require different laboratory methods for processing and nucleic acid extraction (34); bacterial infections are associated with greater pleocytosis and, therefore, higher levels of host background (23); and the analysis of viruses is more tractable given that mNGS (34) commonly detects bacterial reads (e.g., *Pseudomonas aeruginosa* and *Escherichia coli*) as background from skin and reagents (34). As this was not a clinical validation study, we focused on the practical application of mNGS in a defined cohort rather than general diagnostic test performance (21, 23). We adhered to strict practices to minimize contamination, but we did not conduct this research study in a Clinical Laboratory Improvement Amendments (CLIA)-certified laboratory (22), allowing us flexibility in iterative testing and refinement of methods. Because this study was conducted primarily using clinical excess samples, many of which had undergone multiple prior freeze-thaw cycles for clinical testing, it is also possible that some infections were missed due to nucleic acid degradation prior to mNGS, which would be solved if clinical processing for mNGS is standardized (34).

Conclusions. Overall, our results highlight several important benefits of mNGS, including opportunities to reduce dependence on test-specific diagnostics, recover pathogen genomic

data, and potentially offer shorter turnaround times than serology (17). However, our results among subjects with an unknown etiology of disease suggest that the addition of mNGS to standard clinical testing will lead to relatively few additional diagnoses, underscoring the challenge of identifying an etiology in these devastating clinical syndromes. One potential strategy for incorporating mNGS into clinical diagnostic workflows would be wide implementation early in the diagnostic workup to capitalize on the one-step detection of common pathogens, potentially sparing subjects unnecessary tests and reducing overall costs. An alternative would be to reserve this specialized technique for subjects with a high pretest probability of infection (e.g., immunocompromised). Determining how to best utilize mNGS in clinical practice will require evaluation of these factors as well as the cost and logistics of implementation (46, 47). Currently, it is prudent to employ diagnostic mNGS through close communication between clinicians and mNGS experts (18) to evaluate the plausibility of the pathogens identified. This is especially important considering background reads and contamination, the essential limitation that mNGS detects only infections with circulating pathogen nucleic acid, and our still-evolving understanding of mNGS test characteristics. Results from this study will inform ongoing efforts to transition the much-needed and promising technique of mNGS from a research tool to a clinical test used in the routine care of patients with suspected CNS infection.

MATERIALS AND METHODS

Subject enrollment and clinical characterization. The Prospective Encephalitis and Meningitis Study (PEMS) is a prospective cohort study enrolling adults who present to Massachusetts General Hospital (MGH) with confirmed or suspected CNS infection. Adults aged ≥ 18 years with at least one of the following symptoms were eligible for enrollment: (i) altered level of consciousness, (ii) fever, (iii) seizure, (iv) focal neurological findings, (v) electroencephalographic or neuroimaging findings consistent with encephalitis or meningitis, and (vi) refractory headaches. Additionally, enrollment was offered only to patients who had undergone, or planned to undergo, lumbar puncture (LP). Potential participants were referred to the study team by clinicians concerned for encephalitis or meningitis. In parallel, the study team performed queries of the electronic medical record to screen for eligible participants based on the reason for admission and/or CSF results. Further details of screening and enrollment are provided in Text S1 in the supplemental material.

A total of 136 subjects were prospectively enrolled in the PEMS between April 2016 and December 2017, of whom 122 had available CSF for mNGS. For this study, immunocompetent patients with a CSF white blood cell count (WBC) of < 5 cells/ μl ($n = 40$) were excluded as being unlikely to have infectious meningitis or encephalitis; most of these patients had been enrolled prior to undergoing LP (Text S1). Patients with infection due to nosocomial bacteria, or bacteria and fungi that would be challenging to distinguish from common laboratory contamination in mNGS, were also excluded ($n = 14$) (Table S1). Sixty-eight subjects were ultimately included in the mNGS analysis (Fig. 1). This study was approved by the Partners Institutional Review Board under protocol 2015P001388. Further details are in Text S1.

Nucleic acid isolation and standard mNGS. To minimize environmental contamination from viruses studied in the research laboratory, nucleic acid extraction and library construction were performed in an isolated workspace with limited access, extensive decontamination, and strict oversight of supplies, storage areas, and reagents. As a negative control, water and/or CSF from an uninfected patient (negative CSF) was included with each batch starting from nucleic acid isolation. Nucleic acid was extracted from 140 μl of CSF, urine, or plasma stabilized with linear acrylamide using the QIAamp viral RNA minikit (Qiagen). The eluent was split into two fractions for RNA and DNA sequencing. External RNA Controls Consortium (ERCC) spike-in oligonucleotides were added to each fraction; for the RNA fraction, RNA spike-in oligonucleotides were synthesized according to National Institute of Standards and Technology instructions, and for the DNA fraction, cDNA spike-in oligonucleotides were synthesized from RNA templates using random hexamer primers (48). Samples also underwent cDNA synthesis using random hexamer primers and previously described methods (49, 50). Both DNA and RNA libraries underwent tagmentation with the Nextera XT DNA library prep kit (Illumina) and were pooled and sequenced on HiSeq and MiSeq machines using paired-end 100- or 150-bp reads. Methods are outlined in Fig. 1B, and details are in Text S1.

Methods to enhance the detection of pathogen nucleic acid. We attempted two different approaches to enhance sequencing-based detection of pathogen nucleic acid, either separately or together. We first assessed whether enrichment for nonmethylated microbial DNA would improve mNGS yields. We used samples from 12 subjects: 10 with clinically diagnosed DNA virus infections and 2 with clinically diagnosed Lyme disease. Samples underwent methylated DNA depletion (MDD) using the NEBNext microbiome DNA enrichment kit (New England BioLabs), and the enriched fraction was used for DNA library construction as described above (Fig. 4). This kit is designed to remove human host DNA that is methylated and is expected to enrich viral content because no known viral pathogens encode a methyltransferase. We then assessed the efficacy of enrichment for viral nucleic acid by hybrid capture (HC) using a set of probes targeting all viruses known to infect humans (51). We applied HC to 13 RNA and 12 DNA libraries from subjects with clinically diagnosed RNA and DNA virus infections, respectively. Given the observed efficacy of HC, we also

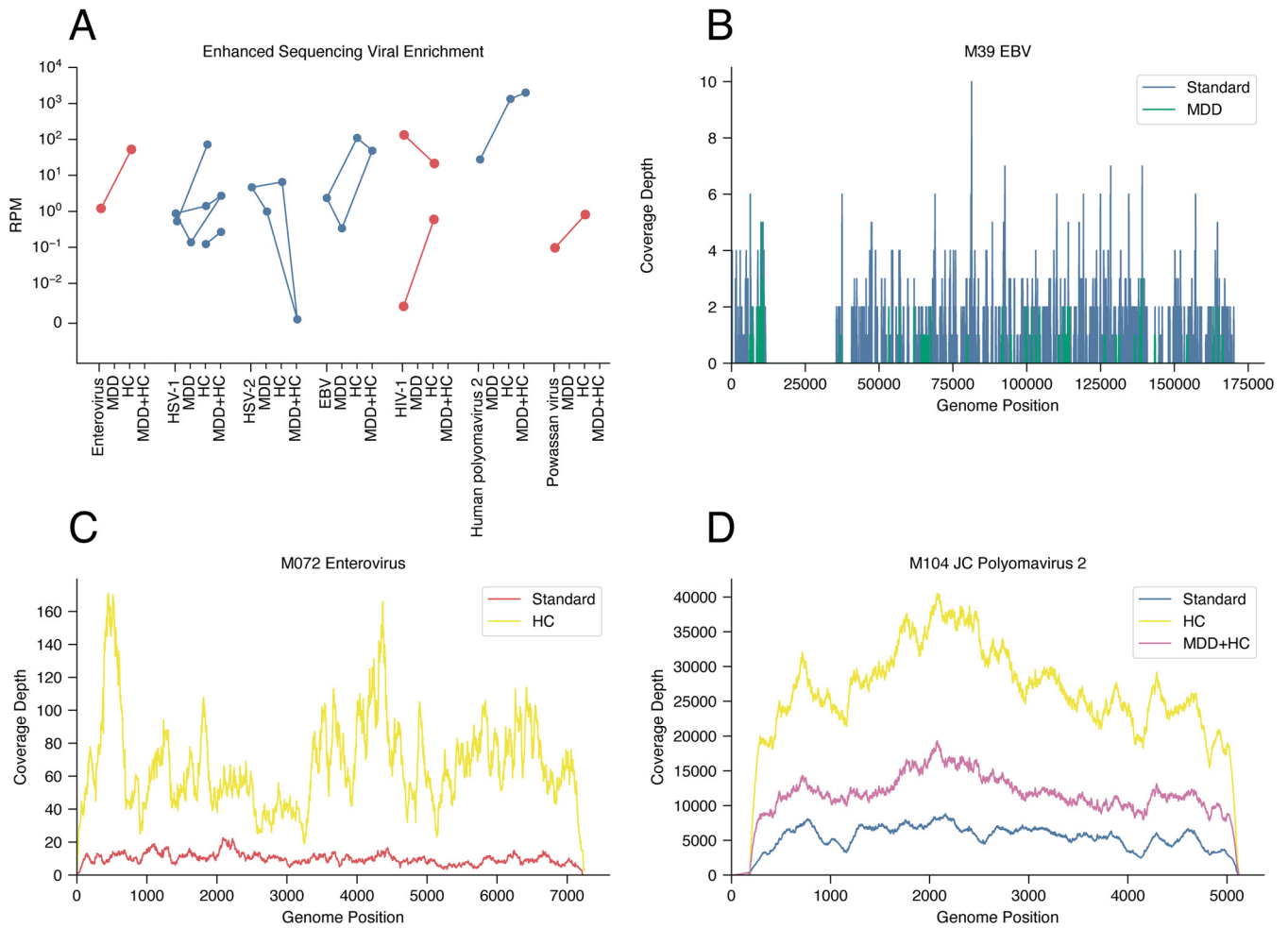


FIG 4 Enhanced methods for mNGS. (A) Comparison of specific viral abundances among the non-computationally depleted reports for HC, MDD, and HC plus MDD for RNA samples (orange) and DNA samples (blue). (B to D) Hybrid capture improved the overall coverage for DNA and RNA viruses such as EBV (B), enterovirus (C), and JC polyomavirus 2 (D). Methylated DNA depletion improved the coverage for some DNA viruses such as JC polyomavirus 2 (D) but not others such as EBV, which utilizes host methylation in its life cycle (B).

applied HC to samples from 20 subjects in the “unknown” group (using the RNA library, DNA library, or both depending on clinical suspicion for a specific pathogen). To perform HC, indexed libraries were pooled into groups of approximately 5 samples per reaction and then underwent hybridization and capture using the SeqCap EZ enrichment kit (Roche), with modifications as described previously (51). HC libraries were pooled and sequenced as described above. Finally, we applied both MDD and HC to a subset of 12 samples from patients with known or suspected DNA virus infection.

Metagenomic analysis pipeline. Illumina sequencing reads were demultiplexed via viral-ngs (57), quality filtered and read trimmed using Trimmomatic (52), and depleted of human reads via a comprehensive KrakenUniq (53) database. The resulting reads were deduplicated and assembled into metagenomic contigs via metaSPAdes (54). Contigs were classified using a cascading BLAST scheme in which unclassified contigs at each stage passed to the next level of more intensive BLAST searches from MegaBLAST and BLASTn to BLASTx (55, 56). Contigs and associated hits derived from water and negative-control samples were aggregated into a contaminant database and used to further deplete the human-depleted reads (Fig. 1C; Fig. S1).

Finally, the human- and contaminant-depleted reads (see Table S4 at <https://figshare.com/articles/dataset/Tables/13266506>) were classified by KrakenUniq using the same comprehensive database as the one described above. Reads classified as potentially human-pathogenic viruses were validated via BLAST, discarding any reads that were not concordantly classified by both methods. The counts of reads per taxon were normalized to sequencing depth as reads per million (RPM) (see Tables S5 and S6 at <https://figshare.com/articles/dataset/Tables/13266506>). Kaiju (58) was run on depleted reads to explore divergent taxon hits, while viral-ngs was used to assemble genomes for a subset of viruses. For mNGS interpretation after computational classification, false-positive species were identified based on broad contamination patterns across all sequencing runs. All true-positive determinations were output from the mNGS classification results directly.

Statistical analyses. Analyses were performed using Student’s *t* test and the Mann-Whitney U test for normally and nonnormally distributed continuous variables, respectively, and using the χ^2 test for categorical variables.

Data availability. Reads after quality control (QC) filtering, trimming, and depletion of human reads via KrakenUniq to a comprehensive database, including the human genome (GRCh38/hg38) and all human sequences from the BLAST NT database, are available in the NCBI Sequence Read Archive (SRA) under accession number [PRJNA668392](https://www.ncbi.nlm.nih.gov/sra/PRJNA668392).

SUPPLEMENTAL MATERIAL

Supplemental material is available online only.

TEXT S1, DOCX file, 0.04 MB.

FIG S1, SVG file, 0.01 MB.

FIG S2, SVG file, 0.1 MB.

FIG S3, SVG file, 0.6 MB.

FIG S4, SVG file, 0.7 MB.

FIG S5, SVG file, 0.2 MB.

FIG S6, SVG file, 0.02 MB.

TABLE S1, XLSX file, 0.01 MB.

TABLE S2, XLSX file, 0.01 MB.

TABLE S3, XLSX file, 0.01 MB.

ACKNOWLEDGMENTS

We are grateful to the patients, their families, and all medical staff involved in their care. We thank Graham McGrath for laboratory support and Janice Stefanski and the microbiology staff at MGH for facilitating sample acquisition.

A.P. was supported by Harvard Catalyst KL2, National Institute of Allergy and Infectious Diseases at the National Institutes of Health (grant number K08AI139348), and the Harvard University Eleanor and Miles Shore Fellowship Program. This work was supported by a gift from the Broad Institute. S.S.M. was supported by the National Institute of Mental Health at the National Institutes of Health (grant number K23MH115812), a developmental award from the Harvard University Center for AIDS Research (HU CFAR NIH/NIAID fund 5P30AI060354-16), and the Harvard University Eleanor and Miles Shore Fellowship Program. The funders of the study had no role in study design, data collection, data analysis, data interpretation, or writing of the report.

P.S. is a cofounder and consultant for Sherlock Biosciences, is a board member of Danaher Corporation, and has equity in both. A.A.A. is a medical director at Karius.

A.P., study design, IRB site principal investigator, data collection, figures, data analyses, data interpretation, and writing. S.S.M., study design, IRB principal investigator, data collection, figures, data analyses, data interpretation, and writing. S.Y., figures, computational analyses, data interpretation, and writing. M.J.L., data collection, figures, and literature search. L.M.F., data collection. D.P., data interpretation and data analyses. G.A., data collection. J.L., study design, data collection, and data interpretation. S.K., data collection and data interpretation. I.H.S., figures, data interpretation, and writing. A.A.A., data interpretation and writing. R.G., study design and data analysis. V.G., data collection. B.O., data collection and data interpretation. K.C.C., figures. J.M.T., writing. C.M.K., data analysis and data interpretation. E.R., study design and data interpretation. M.P.F., study design and data interpretation. M.B.G., study design and data interpretation. T.A.C., study design, data interpretation, and writing. P.S., study design, data interpretation, and writing. A.P., S.S.M., and S.Y. had full access to all the data in the study and final responsibility for the decision to submit for publication.

REFERENCES

1. GBD 2016 Neurology Collaborators. 2019. Global, regional, and national burden of neurological disorders, 1990–2016: a systematic analysis for the Global Burden of Disease Study 2016. *Lancet Neurol* 18:459–480. [https://doi.org/10.1016/S1474-4422\(18\)30499-X](https://doi.org/10.1016/S1474-4422(18)30499-X).
2. McGill F, Griffiths MJ, Bonnett LJ, Geretti AM, Michael BD, Beeching NJ, McKee D, Scarlett P, Hart IJ, Mutton KJ, Jung A, Adan G, Gummery A, Sulaiman WAW, Ennis K, Martin AP, Haycox A, Miller A, Solomon T, UK Meningitis Study Investigators. 2018. Incidence, aetiology, and sequelae of viral meningitis in UK adults: a multicentre prospective observational cohort study. *Lancet Infect Dis* 18:992–1003. [https://doi.org/10.1016/S1473-3099\(18\)30245-7](https://doi.org/10.1016/S1473-3099(18)30245-7).
3. Quist-Paulsen E, Ormaasen V, Kran A-MB, Dunlop O, Ueland PM, Ueland T, Eikeland R, Aukrust P, Nordenmark TH. 2019. Encephalitis and aseptic meningitis: short-term and long-term outcome, quality of life and neuropsychological functioning. *Sci Rep* 9:16158. <https://doi.org/10.1038/s41598-019-52570-2>.

4. Glaser CA, Honarmand S, Anderson LJ, Schnurr DP, Forghani B, Cossen CK, Schuster FL, Christie LJ, Tureen JH. 2006. Beyond viruses: clinical profiles and etiologies associated with encephalitis. *Clin Infect Dis* 43:1565–1577. <https://doi.org/10.1086/509330>.
5. Kupila L, Vuorinen T, Vainionpää R, Hukkanen V, Marttila RJ, Kotilainen P. 2006. Etiology of aseptic meningitis and encephalitis in an adult population. *Neurology* 66:75–80. <https://doi.org/10.1212/01.wnl.0000191407.81333.00>.
6. Neshar L, Hadi CM, Salazar L, Wootton SH, Garey KW, Lasco T, Luce AM, Hasbun R. 2016. Epidemiology of meningitis with a negative CSF gram stain: under-utilization of available diagnostic tests. *Epidemiol Infect* 144:189–197. <https://doi.org/10.1017/S0950268815000850>.
7. Simner PJ, Miller S, Carroll KC. 2018. Understanding the promises and hurdles of metagenomic next-generation sequencing as a diagnostic tool for infectious diseases. *Clin Infect Dis* 66:778–788. <https://doi.org/10.1093/cid/cix881>.
8. Gu W, Miller S, Chiu CY. 2019. Clinical metagenomic next-generation sequencing for pathogen detection. *Annu Rev Pathol* 14:319–338. <https://doi.org/10.1146/annurev-pathmechdis-012418-012751>.
9. Chiu CY, Miller SA. 2019. Clinical metagenomics. *Nat Rev Genet* 20:341–355. <https://doi.org/10.1038/s41576-019-0113-7>.
10. Wilson MR, Naccache SN, Samayoa E, Biagtan M, Bashir H, Yu G, Salamat SM, Somasekar S, Federman S, Miller S, Sokolic R, Garabedian E, Candotti F, Buckley RH, Reed KD, Meyer TL, Serogy CM, Galloway R, Henderson SL, Gern JE, DeRisi JL, Chiu CY. 2014. Actionable diagnosis of neuroleptospirosis by next-generation sequencing. *N Engl J Med* 370:2408–2417. <https://doi.org/10.1056/NEJMoa1401268>.
11. Mongkolrattanothai K, Naccache SN, Bender JM, Samayoa E, Pham E, Yu G, Dien Bard J, Miller S, Aldrovandi G, Chiu CY. 2017. Neurobrucellosis: unexpected answer from metagenomic next-generation sequencing. *J Pediatric Infect Dis Soc* 6:393–398. <https://doi.org/10.1093/jpids/piw066>.
12. Fan S, Ren H, Wei Y, Mao C, Ma Z, Zhang L, Wang L, Ge Y, Li T, Cui L, Wu H, Guan X. 2018. Next-generation sequencing of the cerebrospinal fluid in the diagnosis of neurobrucellosis. *Int J Infect Dis* 67:20–24. <https://doi.org/10.1016/j.ijid.2017.11.028>.
13. Christopeit M, Grundhoff A, Rohde H, Belmar-Campos C, Grzyska U, Fiehler J, Wolschke C, Ayuk F, Kröger N, Fischer N. 2016. Suspected encephalitis with *Candida tropicalis* and *Fusarium* detected by unbiased RNA sequencing. *Ann Hematol* 95:1919–1921. <https://doi.org/10.1007/s00277-016-2770-3>.
14. Wilson MR, Shanbhag NM, Reid MJ, Singhal NS, Gelfand JM, Sample HA, Benkli B, O'Donovan BD, Ali IKM, Keating MK, Dunnebacke TH, Wood MD, Bollen A, DeRisi JL. 2015. Diagnosing *Balamuthia mandrillaris* encephalitis with metagenomic deep sequencing. *Ann Neurol* 78:722–730. <https://doi.org/10.1002/ana.24499>.
15. Wilson MR, Zimmermann LL, Crawford ED, Sample HA, Soni PR, Baker AN, Khan LM, DeRisi JL. 2017. Acute West Nile virus meningoencephalitis diagnosed via metagenomic deep sequencing of cerebrospinal fluid in a renal transplant patient. *Am J Transplant* 17:803–808. <https://doi.org/10.1111/ajt.14058>.
16. Murkey JA, Chew KW, Carlson M, Shannon CL, Sirohi D, Sample HA, Wilson MR, Vespa P, Humphries RM, Miller S, Klausner JD, Chiu CY. 2017. Hepatitis E virus-associated meningoencephalitis in a lung transplant recipient diagnosed by clinical metagenomic sequencing. *Open Forum Infect Dis* 4:ofx121. <https://doi.org/10.1093/ofid/ofx121>.
17. Piantadosi A, Kanjilal S, Ganesh V, Khanna A, Hyle EP, Rosand J, Bold T, Metsky HC, Lemieux J, Leone MJ, Freimark L, Matranga CB, Adams G, McGrath G, Zamirpour S, Telford S, III, Rosenberg E, Cho T, Frosch MP, Goldberg MB, Mukerji SS, Sabeti PC. 2018. Rapid detection of Powassan virus in a patient with encephalitis by metagenomic sequencing. *Clin Infect Dis* 66:789–792. <https://doi.org/10.1093/cid/cix792>.
18. Wilson MR, Sample HA, Zorn KC, Arevalo S, Yu G, Neuhaus J, Federman S, Stryke D, Briggs B, Langelier C, Berger A, Douglas V, Josephson SA, Chow FC, Fulton BD, DeRisi JL, Gelfand JM, Naccache SN, Bender J, Dien Bard J, Murkey J, Carlson M, Vespa PM, Vijayan T, Allyn PR, Campeau S, Humphries RM, Klausner JD, Ganzon CD, Memar F, Ocampo NA, Zimmermann LL, Cohen SH, Polage CR, DeBiasi RL, Haller B, Dallas R, Maron G, Hayden R, Messacar K, Dominguez SR, Miller S, Chiu CY. 2019. Clinical metagenomic sequencing for diagnosis of meningitis and encephalitis. *N Engl J Med* 380:2327–2340. <https://doi.org/10.1056/NEJMoa1803396>.
19. Haston JC, Rostad CA, Jerris RC, Milla SS, McCracken C, Pratt C, Wiley M, Prieto K, Palacios G, Shane AL, McElroy AK. 2020. Prospective cohort study of next-generation sequencing as a diagnostic modality for unexplained encephalitis in children. *J Pediatric Infect Dis Soc* 9:326–333. <https://doi.org/10.1093/jpids/piz032>.
20. Brown JR, Bharucha T, Breuer J. 2018. Encephalitis diagnosis using metagenomics: application of next generation sequencing for undiagnosed cases. *J Infect* 76:225–240. <https://doi.org/10.1016/j.jinf.2017.12.014>.
21. Schlager R, Chiu CY, Miller S, Procop GW, Weinstock G, Professional Practice Committee and Committee on Laboratory Practices of the American Society for Microbiology, Microbiology Resource Committee of the College of American Pathologists. 2017. Validation of metagenomic next-generation sequencing tests for universal pathogen detection. *Arch Pathol Lab Med* 141:776–786. <https://doi.org/10.5858/arpa.2016-0539-RA>.
22. Blauwkamp TA, Thair S, Rosen MJ, Blair L, Lindner MS, Vilfan ID, Kawli T, Christians FC, Venkatasubrahmanyam S, Wall GD, Cheung A, Rogers ZN, Meshulam-Simon G, Huijse L, Balakrishnan S, Quinn JV, Hollemon D, Hong DK, Vaughn ML, Kertesz M, Bercovici S, Wilber JC, Yang S. 2019. Analytical and clinical validation of a microbial cell-free DNA sequencing test for infectious disease. *Nat Microbiol* 4:663–674. <https://doi.org/10.1038/s41564-018-0349-6>.
23. Miller S, Naccache SN, Samayoa E, Messacar K, Arevalo S, Federman S, Stryke D, Pham E, Fung B, Bolosky WJ, Ingebrigtsen D, Lorizio W, Paff SM, Leake JA, Pesano R, DeBiasi R, Dominguez S, Chiu CY. 2019. Laboratory validation of a clinical metagenomic sequencing assay for pathogen detection in cerebrospinal fluid. *Genome Res* 29:831–842. <https://doi.org/10.1101/gr.238170.118>.
24. Glaser CA, Gilliam S, Schnurr D, Forghani B, Honarmand S, Khetsuriani N, Fischer M, Cossen CK, Anderson LJ. 2003. In search of encephalitis etiologies: diagnostic challenges in the California Encephalitis Project, 1998–2000. *Clin Infect Dis* 36:731–742. <https://doi.org/10.1086/367841>.
25. Vora NM, Holman RC, Mehal JM, Steiner CA, Blanton J, Sejvar J. 2014. Burden of encephalitis-associated hospitalizations in the United States, 1998–2010. *Neurology* 82:443–451. <https://doi.org/10.1212/WNL.000000000000086>.
26. Gilden DH, Mahalingam R, Cohrs RJ, Tyler KL. 2007. Herpesvirus infections of the nervous system. *Nat Clin Pract Neurol* 3:82–94. <https://doi.org/10.1038/ncpneuro0401>.
27. Aburakawa Y, Katayama T, Saito T, Sawada J, Suzutani T, Aizawa H, Hasebe N. 2017. Limbic encephalitis associated with human herpesvirus-7 (HHV-7) in an immunocompetent adult: the first reported case in Japan. *Intern Med* 56:1919–1923. <https://doi.org/10.2169/internalmedicine.56.8185>.
28. Riva N, Franconi I, Meschiari M, Franceschini E, Puzzolante C, Cuomo G, Bianchi A, Cavalleri F, Genovese M, Mussini C. 2017. Acute human herpes virus 7 (HHV-7) encephalitis in an immunocompetent adult patient: a case report and review of literature. *Infection* 45:385–388. <https://doi.org/10.1007/s15010-017-1014-3>.
29. Parra M, Alcalá A, Amorós C, Baeza A, Galiana A, Tarragó D, García-Quesada MA, Sánchez-Hellín V. 2017. Encephalitis associated with human herpesvirus-7 infection in an immunocompetent adult. *Virology* 14:97. <https://doi.org/10.1186/s12985-017-0764-y>.
30. Asplund M, Kjartansdóttir KR, Møllerup S, Vinner L, Fridholm H, Herrera JAR, Friis-Nielsen J, Hansen TA, Jensen RH, Nielsen IB, Richter SR, Rey-Iglesia A, Matey-Hernandez ML, Alquezar-Planas DE, Olsen PVS, Sicheritz-Pontén T, Willerslev E, Lund O, Brunak S, Mourier T, Nielsen LP, Izarzugaza JMG, Hansen AJ. 2019. Contaminating viral sequences in high-throughput sequencing viromics: a linkage study of 700 sequencing libraries. *Clin Microbiol Infect* 25:1277–1285. <https://doi.org/10.1016/j.cmi.2019.04.028>.
31. Shi M, Lin X-D, Tian J-H, Chen L-J, Chen X, Li C-X, Qin X-C, Li J, Cao J-P, Eden J-S, Buchmann J, Wang W, Xu J, Holmes EC, Zhang Y-Z. 2016. Redefining the invertebrate RNA virosphere. *Nature* 540:539–543. <https://doi.org/10.1038/nature20167>.
32. Vareil M, Wille H, Kassab S, Le-Cornec C, Puges M, Desclaux A, Lafon ME, Tumiotto C, Cazanave C, Neau D. 2018. Clinical and biological features of enteroviral meningitis among adults and children and factors associated with severity and length of stay. *J Clin Virol* 104:56–60. <https://doi.org/10.1016/j.jcv.2018.04.017>.
33. Thoendel M, Jerald PR, Greenwood-Quaintance KE, Yao JZ, Chia N, Hanssen AD, Abdel MP, Patel R. 2016. Comparison of microbial DNA enrichment tools for metagenomic whole genome sequencing. *J Microbiol Methods* 127:141–145. <https://doi.org/10.1016/j.mimet.2016.05.022>.
34. Simner PJ, Miller HB, Breitwieser FP, Pinilla Monsalve G, Pardo CA, Salzberg SL, Sears CL, Thomas DL, Eberhart CG, Carroll KC. 2018. Development and optimization of metagenomic next-generation sequencing methods for cerebrospinal fluid diagnostics. *J Clin Microbiol* 56:e00472-18. <https://doi.org/10.1128/JCM.00472-18>.
35. Hasan MR, Rawat A, Tang P, Jithesh PV, Thomas E, Tan R, Tilley P. 2016. Depletion of human DNA in spiked clinical specimens for improvement of sensitivity of pathogen detection by next-generation sequencing. *J Clin Microbiol* 54:919–927. <https://doi.org/10.1128/JCM.03050-15>.

36. Charalampous T, Kay GL, Richardson H, Aydin A, Baldan R, Jeanes C, Rae D, Grundy S, Turner DJ, Wain J, Leggett RM, Livermore DM, O'Grady J. 2019. Nanopore metagenomics enables rapid clinical diagnosis of bacterial lower respiratory infection. *Nat Biotechnol* 37:783–792. <https://doi.org/10.1038/s41587-019-0156-5>.
37. Rosenberg R, Lindsey NP, Fischer M, Gregory CJ, Hinckley AF, Mead PS, Paz-Bailey G, Waterman SH, Drexler NA, Kersh GJ, Hooks H, Partridge SK, Visser SN, Beard CB, Petersen LR. 2018. Vital signs: trends in reported vectorborne disease cases—United States and territories, 2004–2016. *MMWR Morb Mortal Wkly Rep* 67:496–501. <https://doi.org/10.15585/mmwr.mm6717e1>.
38. Bennett JE, Dolin R, Blaser MJ (ed). 2014. *Mandell, Douglas, and Bennett's principles and practice of infectious diseases*, 8th ed. Elsevier Health Sciences, Philadelphia, PA.
39. Dahlgren FS, Mandel EJ, Krebs JW, Massung RF, McQuiston JH. 2011. Increasing incidence of Ehrlichia chaffeensis and Anaplasma phagocytophilum in the United States, 2000–2007. *Am J Trop Med Hyg* 85:124–131. <https://doi.org/10.4269/ajtmh.2011.10-0613>.
40. Dunn BE, Monson TP, Dumler JS, Morris CC, Westbrook AB, Duncan JL, Dawson JE, Sims KG, Anderson BE. 1992. Identification of Ehrlichia chaffeensis morulae in cerebrospinal fluid mononuclear cells. *J Clin Microbiol* 30:2207–2210. <https://doi.org/10.1128/jcm.30.8.2207-2210.1992>.
41. Lyons JL, Schaefer PW, Cho TA, Azar MM. 2017. Case 34-2017. A 76-year-old man with fever, weight loss, and weakness. *N Engl J Med* 377:1878–1886. <https://doi.org/10.1056/NEJMcpc1707557>.
42. Huang C, Slater B, Rudd R, Parchuri N, Hull R, Dupuis M, Hindenburg A. 2002. First isolation of West Nile virus from a patient with encephalitis in the United States. *Emerg Infect Dis* 8:1367–1371. <https://doi.org/10.3201/eid0812.020532>.
43. Morjaria S, Arguello E, Taur Y, Sepkowitz K, Hatzoglou V, Nemade A, Rosenblum M, Cavalcanti MS, Palomba ML, Kaltsas A. 2015. West Nile virus central nervous system infection in patients treated with rituximab: implications for diagnosis and prognosis, with a review of literature. *Open Forum Infect Dis* 2:ofv136. <https://doi.org/10.1093/ofid/ofv136>.
44. Levi ME, Quan D, Ho JT, Kleinschmidt-Demasters BK, Tyler KL, Grazia TJ. 2010. Impact of rituximab-associated B-cell defects on West Nile virus meningoencephalitis in solid organ transplant recipients. *Clin Transplant* 24:223–228. <https://doi.org/10.1111/j.1399-0012.2009.01044.x>.
45. Torres SD, Jia DT, Schorr EM, Park BL, Boubour A, Boehme A, Ankam JV, Gofshsteyn JS, Tyshkov C, Green DA, Vargas W, Zucker J, Yeshokumar AK, Thakur KT. 2020. Central nervous system (CNS) enterovirus infections: a single center retrospective study on clinical features, diagnostic studies, and outcome. *J Neurovirol* 26:14–22. <https://doi.org/10.1007/s13365-019-00784-5>.
46. Greninger AL. 2018. The challenge of diagnostic metagenomics. *Expert Rev Mol Diagn* 18:605–615. <https://doi.org/10.1080/14737159.2018.1487292>.
47. Hogan CA, Yang S, Garner OB, Green DA, Gomez CA, Dien Bard J, Pinsky BA, Banaei N. 2021. Clinical impact of metagenomic next-generation sequencing of plasma cell-free DNA for the diagnosis of infectious diseases: a multi-center retrospective cohort study. *Clin Infect Dis* 72:239–245. <https://doi.org/10.1093/cid/ciaa035>.
48. Fasolka MJ. 2017. DNA sequence library for external RNA controls. National Institute of Standards and Technology, Gaithersburg, MD.
49. Gire SK, Goba A, Andersen KG, Sealfon RSG, Park DJ, Kanneh L, Jalloh S, Momoh M, Fullah M, Dudas G, Wohl S, Moses LM, Yozwiak NL, Winnicki S, Matranga CB, Malboeuf CM, Qu J, Gladden AD, Schaffner SF, Yang X, Jiang P-P, Nekoui M, Colubri A, Coomber MR, Fonnies M, Moigboi A, Gbakie M, Kamara FK, Tucker V, Konuwa E, Saffa S, Sellu J, Jalloh AA, Kovoma A, Koninga J, Mustapha I, Kargbo K, Foday M, Yillah M, Kanneh F, Robey W, Massally JLB, Chapman SB, Bochicchio J, Murphy C, Nusbaum C, Young S, Birren BW, Grant DS, Scheffelin JS, Lander ES, Happi C, Gevaio SM, Gnirke A, Rambaut A, Garry RF, Khan SH, Sabeti PC. 2014. Genomic surveillance elucidates Ebola virus origin and transmission during the 2014 outbreak. *Science* 345:1369–1372. <https://doi.org/10.1126/science.1259657>.
50. Matranga CB, Andersen KG, Winnicki S, Busby M, Gladden AD, Tewhey R, Stremlau M, Berlin A, Gire SK, England E, Moses LM, Mikkelsen TS, Oda I, Ehiane PE, Folarin O, Goba A, Kahn SH, Grant DS, Honko A, Hensley L, Happi C, Garry RF, Malboeuf CM, Birren BW, Gnirke A, Levin JZ, Sabeti PC. 2014. Enhanced methods for unbiased deep sequencing of Lassa and Ebola RNA viruses from clinical and biological samples. *Genome Biol* 15:519. <https://doi.org/10.1186/s13059-014-0519-7>.
51. Metsky HC, Siddle KJ, Gladden-Young A, Qu J, Yang DK, Brehio P, Goldfarb A, Piantadosi A, Wohl S, Carter A, Lin AE, Barnes KG, Tully DC, Corleis B, Hennigan S, Barbosa-Lima G, Vieira YR, Paul LM, Tan AL, Garcia KF, Parham LA, Oda I, Eromon P, Folarin OA, Goba A, Viral Hemorrhagic Fever Consortium, Simon-Lorière E, Hensley L, Balmaseda A, Harris E, Kwon DS, Allen TM, Runstadler JA, Smole S, Bozza FA, Souza TML, Isern S, Michael SF, Lorenzana I, Gehrke L, Bosch I, Ebel G, Grant DS, Happi CT, Park DJ, Gnirke A, Sabeti PC, Matranga CB. 2019. Capturing sequence diversity in metagenomes with comprehensive and scalable probe design. *Nat Biotechnol* 37:160–168. <https://doi.org/10.1038/s41587-018-0006-x>.
52. Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
53. Breitwieser FP, Baker DN, Salzberg SL. 2018. KrakenUniq: confident and fast metagenomics classification using unique k-mer counts. *Genome Biol* 19:198. <https://doi.org/10.1186/s13059-018-1568-0>.
54. Nurk S, Meleshko D, Korobeynikov A, Pevzner PA. 2017. metaSPAdes: a new versatile metagenomic assembler. *Genome Res* 27:824–834. <https://doi.org/10.1101/gr.213959.116>.
55. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. *J Mol Biol* 215:403–410. [https://doi.org/10.1016/S0022-2836\(05\)80360-2](https://doi.org/10.1016/S0022-2836(05)80360-2).
56. Morgulis A, Coulouris G, Raytselis Y, Madden TL, Agarwala R, Schäffer AA. 2008. Database indexing for production MegaBLAST searches. *Bioinformatics* 24:1757–1764. <https://doi.org/10.1093/bioinformatics/btn322>.
57. Tomkins-Tinch C, Park DJ, Ye S, et al. 2016. viral-ngs v1.25.0. <https://github.com/broadinstitute/viral-ngs/releases/tag/v1.25.0>.
58. Menzel P, Ng KL, Krogh A. 2016. Fast and sensitive taxonomic classification for metagenomics with Kaiju. *Nat Commun* 7:11257.